

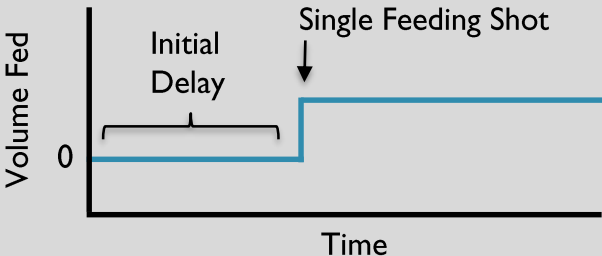
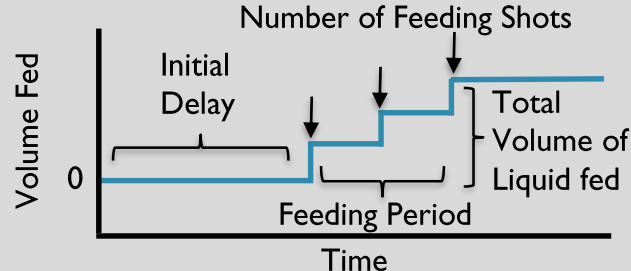
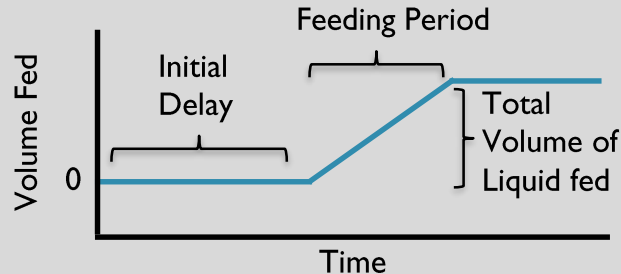
The image shows four Erlenmeyer flasks arranged in a row, each containing a bright blue liquid. Each flask is fitted with a black, rectangular automated injection system (LIS) mounted on its neck. The flasks are graduated with volume markings (100, 150, 200, 250 ml). The background is a soft-focus laboratory setting with a person in a white lab coat visible on the right. A semi-transparent white banner is overlaid across the middle of the image, containing the title and subtitle.

Liquid Injection System (LIS)

Automated Feeding of Liquids in Shake Flasks

Various predefined feeding profiles allow users to quickly create individual feeding strategies for their shake flask cultures.

LIS Feeding Profiles

	Feeding Profile	Feeding Parameters	Description
Single Shot		<ul style="list-style-type: none"> – Initial Delay [min] – Volume of Feeding Shot [μl] 	Allows you to feed a certain volume to your culture after a defined initial delay. This profile could be used for e.g. an induction with IPTG.
Multi Shot		<ul style="list-style-type: none"> – Initial Delay [min] – Feeding Period [min] – Total Volume that should be fed [μl] – Number of Feeding Shots during Feeding Period 	Allows you to feed a certain volume in a defined time period. The liquid will be distributed evenly among a defined number of shots. This profile could be used for e.g. a fed batch experiment.
Constant		<ul style="list-style-type: none"> – Initial Delay [min] – Feeding Period [min] – Total Volume that should be fed [μl] 	Allows you to feed a certain volume in a defined time period. The liquid will be distributed linearly over time. This profile could be used for e.g. a fed batch experiment.

Additional feeding profiles are available in the LIS Software.

LIS is compatible with various substances such as sugars, alcohols as well as acids or bases.

LIS – Compatible Substances

Substance	Compatibility of substance with LIS			
	Unlimited	Under most circumstances	Can not be used	Not tested
Sugar Solutions (e.g. up to 50% Glc)	×			
Viscous Solutions (e.g. up to 75% Glycerol)	×			
Aqueous Solutions (e.g. IPTG, Media, Acids, Bases, Buffer)	×			
Alcohols (e.g. 50% Ethanol, 75% Methanol)		×		
Antifoam		×		
Organic Solvents				×
Phages				×
Suspensions				×
Solid Materials			×	

* Based on the alcohol, concentration and temperature, evaporation may influence the accuracy

** Accuracy can vary for different types of antifoams

LIS is compatible with all common laboratory shakers and shaking conditions.

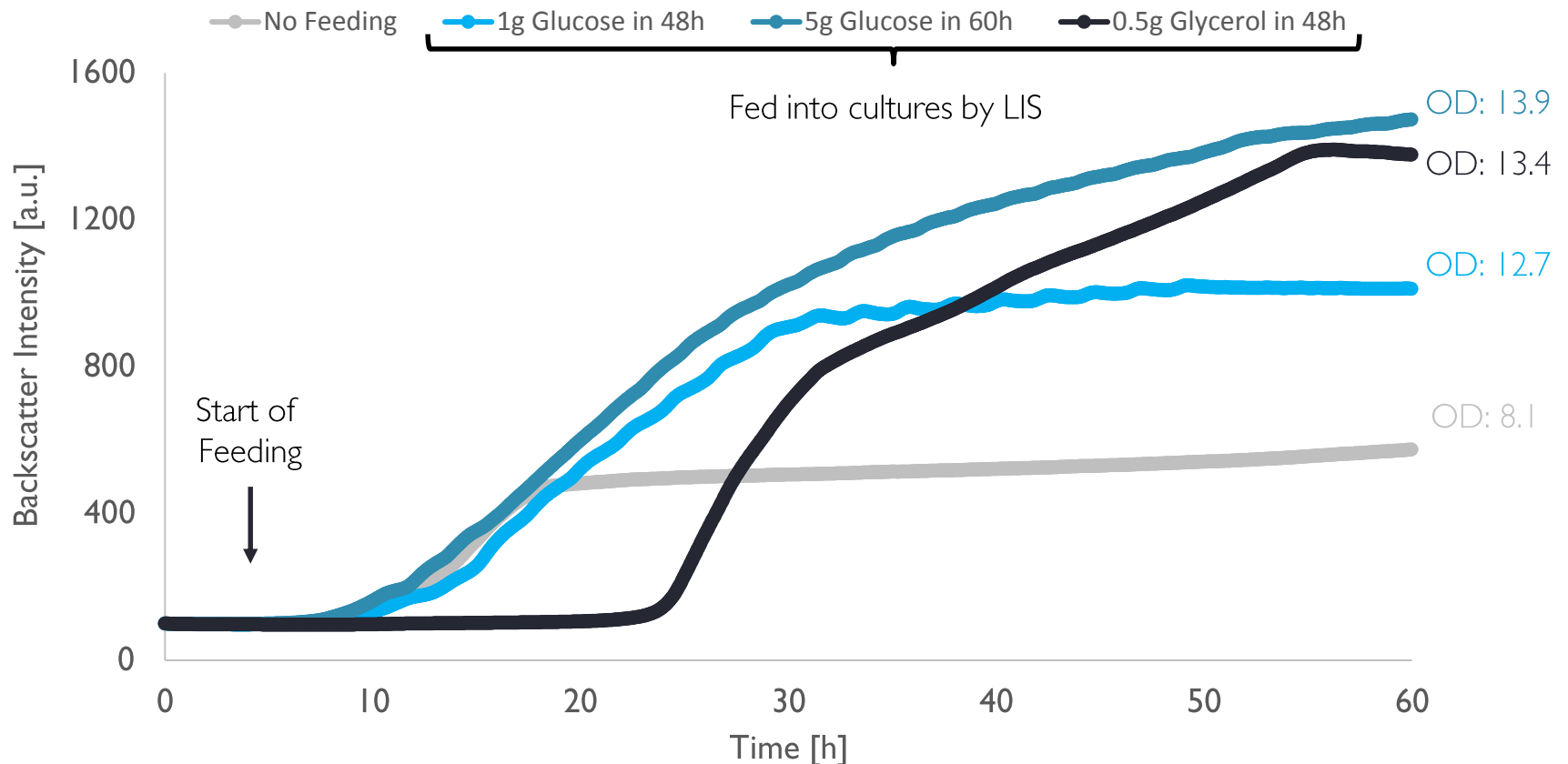
LIS – Compatible Maximal Shaking Conditions

Shaking diameter	Flasks fixed on tray with	Set-Up	Shake Flask Size (Total Volume)				
			100ml	250ml	500ml	1000ml	2000ml
25mm	Clamps	Shake Flask only	250	300	250	250	250
		Shake Flask + "New" CGQ Sensor	250	300	300	300	250
		Shake Flask + "Old" CGQ Sensor		300	300	300	250
	Sticky Stuff	Without CGQ Sensor	200	200	200	250	250
50mm	Clamps	Shake Flask only	250	250	225	225	200
		Shake Flask + "New" CGQ Sensor	200	250	250	225	200
		Shake Flask + "Old" CGQ Sensor		250	225	225	200
	Sticky Stuff	Without CGQ Sensor	200	200	200	250	250

Tests were performed with a flask filling volume of 10% and 20ml filling volume of the LIS cartridge.
All data is shown in rounds per minute (rpm).

LIS is the first easy-to-use technology allowing for fed-batch experiments in shake flask cultures.

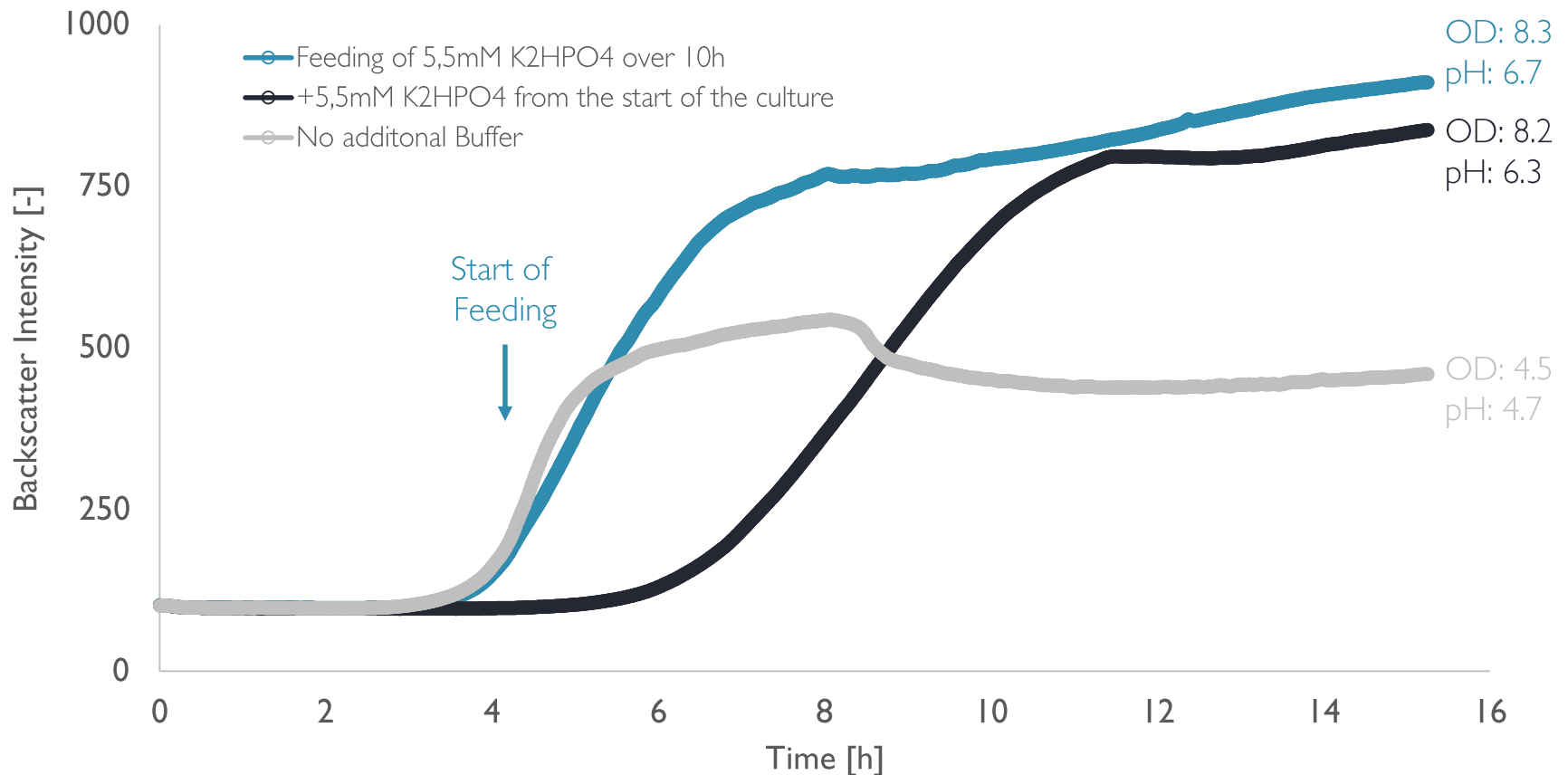
Exemplary Use Case LIS (1/2) – Fed-Batch



S. cerevisiae cultures were grown on YPD media. Cultures were performed in 250ml shake flasks with a filling volume of 10% at 30°C and shaken at 250rpm. Additional carbon source was fed to three culture using the LIS technology. Either Glucose or Glycerol was fed to the cultures over a time period of 48 or 60h by using a constant feeding profile. Biomass of the cultures was monitored online by the CGQ technology (aquila biolabs). OD was measured offline at the end of the cultures. OD = Optical Density

LIS can be used to control pH drifts of *E.coli* shake flask cultures by automatically feeding buffer to the culture.

Exemplary Use Case LIS (2/2) – pH Regulation



E. coli cultures were grown on LB media + 2% Glucose. Cultures were performed in 250ml shake flasks with a filling volume of 10% at 37°C and shaken at 250rpm. Additional buffer was fed to one culture using the LIS technology. 2ml of 5,5mM K₂HPO₄ were fed to the culture over a time period of 10h by using a constant feeding profile. Biomass of the cultures was monitored online by the CGQ technology (aquila biolabs). OD and pH was measured offline at the end of the cultures. OD = Optical Density